Diseases of Banksia woodlands on the Bassendean and Spearwood Dune Systems

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Current knowledge

Diseases of Banksia woodlands have been a neglected area of plant pathology. Of the c 250 000 publications on plant diseases abstracted in the Review of Plant Pathology since 1922, only about 30 refer to diseases of Banksia. Only 6% of these 30 publications refer to diseases of Banksia in woodlands compared with 55% for forest and 39% for Banksia species used in floriculture. Observations on the impact and spread of Phytophthora cinnamomi by Podger (1972) and Havel (1979) are the only published account of disease on Banksia woodlands of the Bassendean Dune system. Nevertheless, despite the lack of published information, disease is an important factor affecting the ecology of Banksia communities.

Phytophthora species have been the most frequent cause of disease of Banksia (73% of the 30 publications) followed by wood rots (12%), leaf spots (9%) and Cylindrocladium scoparium (6%). The leaf spot Asterina systema-solare (Shivas in press), wood rots caused by Armillaria luteobubalina (Shearer & Tippett 1988), Ganoderma, Polyporus, Poria and Stereum (Hilton collection, WA Herbarium) and the canker pathogen Botryosphaeria ribis (Shivas in press) have been recorded on Banksia species occurring on the Bassendean Dunes. However the most destructive impact on the Banksia community of the Bassendean Dune system is disease caused by Phytophthora species, especially P. cinnamomi (Podger 1968, 1972).

Phytophthora cinnamomi is distributed widely in Banksia woodlands of the coastal plain killing most of the overstorey and shrub layers in affected areas (Podger 1972). Incidence of disease is greatest south of Perth, decreasing north of Wanneroo (Havel 1979). The Moore River National Park is the most northerly known occurrence of P. cinnamomi on the coastal plain. Geographically restricted and susceptible B. laricina is being killed in affected areas in this park. The incidence of P. cinnamomi on the Spearwood Dunes is much less than on the Bassendean Dunes (Podger 1968), even though plant species are susceptible and disturbance from human activity is high (Havel 1979).

Phytophthora cinnamomi is an introduced soil-borne fungus belonging to the Oomycetes or "water moulds". As the name "water mould" suggests, the life cycle of P. cinnamomi depends on moist conditions that favour survival, sporulation and dispersal of the fungus, and host infection. Warm, moist conditions and interactions with soil microflora favour vegetative production of sporangia and thick walled chlamydospores from mycelial strands in the soil or host tissue. Interaction of mycelium of different mating types may produce thick-walled sexual oospores. However reproduction in soil is mainly by the asexual

sporangium-zoospore cycle which produces large numbers of infectious spores when conditions are favourable.

Sporangia release motile zoospores in free water. Zoospores can swim over short distances in water, but are mainly dispersed over large distances in flowing water or in infected moist soil moved by human activity. Zoospores in moist soil are chemotactically attracted to root surfaces where they germinate to produce germ tubes that penetrate roots. Infection by *P. cinnamomi* is probably favoured by the thin bark and proliferation of rootlets associated with the specialized proteoid roots of the *Banksia* species occurring on the Bassendean Dunes. The fungus actively grows through root systems or is passively dispersed in infected roots transported in soil. Root to root contact facilitates mycelial growth between root systems and initiation of new infections.

The pathogen infects at least 1000 species of known hosts from taxonomically diverse families (Zentmyer 1980). The families Epacridaceae, Myrtaceae and Proteaceae, important components of *Banksia* woodlands, contain many susceptible species.

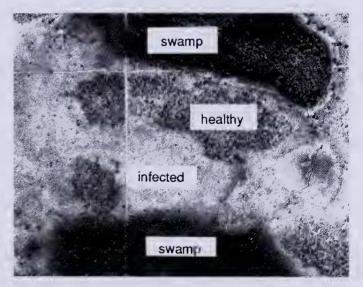


Figure 1 Destruction of *Banksia* woodland following infection by *Phytophthora cinnamomi* (light grey) compared with remnants of healthy woodland (dark grey) on a gently sloping Bassendean Dune between two swamp systems at Gnangara on October 1964. The infected area was healthy woodland prior to 1953. Scale 1:12 500.

Current research

Following Podger's (1968) initial monitoring of sites from Ravenswood to Moore River, little research has been done on the occurrence of *P. cinnamomi* in *Banksia* woodlands on the coastal plain. Investigations are now in progress to determine the factors influencing disease development, impact and methods of control of *P. cinnamomi* and other *Phytophthora* species in *Banksia* woodlands.

Disease Development

The spread of P. cinnamomi in 132 ha of Bassendean Dunes at Gnangara has been mapped from aerial photographs. The area includes 97 ha of Banksia woodland and 35 ha of ephemeral swamp. Four small patches of dead vegetation totalling 0.15 ha occurred in 1942 alongside tracks that radiated from nearby strawberry farms. The area of infected woodland and swamp had increased to 55 ha by 1959, because of expansion of original infections, new infections from a nearby farm and contamination of the swamp system with associated destruction of low lying Banksia woodland (Fig. 1). These disease fronts expanded at 1.0 m yr⁻¹ and 67 ha was infected by 1974, increasing to 82 ha (63%) in 1988. Slope or depth to water table did not appear to influence the rate of spread. However the rate of spread at Gnangara was slower than a mean downslope spread of 8 m yr⁻¹ observed by Podger (1968) on a gently sloping dune of Gavin sand in the Bassendean system near North Dandalup.

The pattern of disease development at Gnangara reflects the ability of P. cinnamomi to exploit various mechanisms of spread. Disturbance associated with market gardening, roads, tracks and off-road driving have resulted in the dispersal of P. cinnamomi in infected soil, and is responsible for the widespread distribution of the pathogen throughout the coastal plain. Active and passive dispersal of zoospores in free water contributes to spread within an area and results in the contamination of swampy areas. Zoospore dispersal in coarse-textured sands may also be assisted by movement of the water table or by lateral drainage from perched layers of saturated soil over clay or iron hardpan. We have recovered the fungus from groundwater at 3 and 5 m below the soil surface in affected Banksia woodland on Gavin sand near Hamel and south of Busselton. Growth of P. cinnamomi in roots of susceptible hosts ensures continued spread through summer, even though activity of the fungus in dry soil ceases. For example, P. cinnamomi can grow up to 1 cm day 1 in roots of susceptible B. grandis in summer when temperatures are optimal for fungal growth (Shearer et al 1987).

The destructiveness and persistence of *P. cinnamomi* in *Banksia* woodlands is partly determined by the ability of the pathogen to survive the dry soil environment over summer and resume activity when moist conditions return. The fungus survived throughout the year in soil sampled from a depth of a metre in an affected *Banksia* woodland on Gavin sand south of Busselton. Recovery rates at depth from this site were often higher than those obtained from a high impact site in the jarrah forest

Infected host tissue provides a buffered environment for P. cinnamomi survival during dry conditions. For example, the fungus survived summer in 65% of colonized pine plugs buried at 30 cm in an affected area at Gnangara, even though soil moisture at this depth decreased to 0.6% in February.

Impact

Phytophthora cinnamomi infection destroys the structure and diversity of Banksia woodland. The dominant overstorey of B. attenuata, B. ilicifolia and B. menziesii is killed and only scattered Eucalyptus todtiana and Nuytsia floribunda remain in affected areas. Many understorey shrub species are similarly affected. Species richness in 64 m^2 quadrats decreased from 56 species

in healthy woodland to 41 species in an affected area. Biomass can be reduced by up to 90% following infection (Fig. 1). Despite the impact of $P.\ cinnamomi$ on Banksia woodland, information is lacking on the long term structural and floristic changes in affected areas.

Other Phytophthora species

Phytophthora citricola. P. cryptogea (A₁), P. megasperma var. megasperma and P. megasperma var. sojae have been isolated from dying vegetation on Bassendean Dunes north of the Moore River. Many of the affected areas were low-lying and seasonally inundated or received off-road drainage. The susceptibility of native vegetation to these Phytophthora species needs to be determined before their relative significance to the health of native plant communities can be accurately evaluated (Shearer et al 1988).

Control

Eradication of *P. cinnamomi* from spot infections by Ridomil and fumigation with formaldehyde is being assessed in Jandakot sands of the Bassendean Dunes at Gnangara with promising results. In addition, the systemic fungicide phosphorous acid has arrested lesion extension in *B. grandis*. Evaluation of these control methods is continuing.

Conclusions

Systematic surveys of *Banksia* woodlands in southwestern Australia are needed to address the lack of information on diseases of *Banksia*.

Phytophthora cinnamomi and other Phytophthora species are major factors affecting the ecology and management of the diverse, but susceptible, Banksia communities on leached sands. Information is lacking on the specific requirements for pathogen survival, sporulation and spread as well as host infection and susceptibility in sandy soils. Such information is essential for the development of hazard and risk systems to minimize introduction and spread of Phytophthora species.

Knowledge of the diversity of *Banksia* woodlands, similar to the site-vegetation classification of Havel (1979), is needed in the development and application of hazard and risk systems. Long term effects of *Phytophthora* spp on diversity clearly needs to be quantified.

An understanding of the low incidence of *P. cinnamomi* on Spearwood Dunes may provide clues for the control of the disease. Control strategies must be developed and applied to prevent spread and intensification of disease favoured by disturbance caused by increasing urbanization and sand mining.

References

Havel J J 1979 Vegetation: Natural factors and human activity. In: Western Landscapes (ed J Gentilli). Univ W Aust 122-152.

Podger F D 1968 Aetiology of jarrah dieback. A disease of dry sclerophyll *Eucalyptus* marginata Sm. forests in Western Australia, M Sc thesis, Univ Melbourne.

Podger F D 1972 Phytophthora cinnamomi, a cause of lethal disease in indigenous plant communities in Western Australia. Phytopathology 62: 972-981.

Shearer B L & Tippett J T 1988 Distribution and impact of Armillaria luteobubalina in the Eucalyptus marginata forest of south-western Australia. Aust J Bot 36: 433-445.

Shearer B L, Shea S R & Deegan P M 1987 Temperature-growth relationships of Phytophthora cinnamomi in the secondary phloem of Banksia grandis and Eucalyptus marginata. Phytopathology 77: 661-665.

Shearer B L, Michaelsen B J & Somerford P J 1988 Effects of isolate and time of inoculation on invasion of secondary phloem of Eucalyptus spp. and Banksia grandis by Phytophthora spp. Plant Disease 72: 121-126.

Shivas R G (in press) Fungal and bacterial diseases in Western Australia. J R Soc W A.

Zentmyer G A 1980 Phytophthora cınnamomi and the diseases it causes. Monograph 10. American Phytopathological Soc, St Paul.